

Cirrhosis in children from peanut meal contaminated by aflatoxin¹

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The most toxic of the metabolites of *Aspergillus flavus* known to grow on peanuts is aflatoxin B₁ (1). It affects the young of many species of mammals and causes hepatic necrosis followed by bile duct proliferation (2). There is, however, no information reported so far on the toxic manifestations of its compound in human beings. With a peak incidence at 5 years, Indian childhood cirrhosis occurs widely on this subcontinent, causing vague gastrointestinal symptoms with anorexia followed by firm hepatomegaly with a characteristic leafy border that may proceed to jaundice, ascites, and hepatic coma (4). In a separate communication from a laboratory (5), the presence of aflatoxin (77% of the urine samples of cirrhotic children as seen on thin-layer chromatoplates) is reported (6). Chromatographic identification was confirmed by the chemical test (Andrèlles and Reid (7)).

Crude peanut oil and parboiled rice obtained from the homes of these children and random market samples that we did revealed significant aflatoxin contamination of 50 to 90 µg per kg (8) (data).

Based on these observations, the possibility of involvement of aflatoxin in the etiology of Indian childhood cirrhosis is suggested.

Materials and methods

In January 1969, 20 children (13 boys and 7 girls) aged 1 to 5 years, respectively, and affected by Indian childhood cirrhosis, were selected for the study. They were all from a low-income group in Mysore town, as when compared with the general population of the town, the incidence of cirrhosis is reported to be higher (9). Informed consent was obtained from the parents.

Subsequent to the selection of the children, a body fluid sample was obtained from each child during the previous 2 months. The following signs and symptoms, which were observed during the illness, were noted: anorexia, vomiting, weight loss, abdominal distention, hepatomegaly, ascites, jaundice, and hepatic coma. The clinical course of the disease was as follows: 13 children were hospitalized in the

ward was found to be 100% µg per kg. With this information, all the children who were known to have consumed the contaminated peanut meal were selected for the study. The study group comprised 20 children aged 1.5 years to 5 years, all of whom had consumed 1 to 2 oz of the toxic material daily for periods varying from 7 days to 3 months. All were cases of protein-calorie malnutrition with the exception of one normal child and one who had nephrosis. Detailed dietary history was obtained with particular reference to crude peanut oil, peanuts, and parboiled rice. There was no history of the consumption of these food items, as these children belonged to the lowest income group and their diet was comprised mainly of ragi (*Mesquite coracaria*), a little milk, and vegetables. In our weekly clinic, the children were carefully examined, and records of hepatomegaly and other laboratory investigations were maintained. Repeated liver biopsies were taken every 2 to 3 months, and it is the practice in this clinic to take a liver biopsy after the first examination. In many children, three to four liver biopsies were done at 2- to 3-month intervals. In others, because they failed to attend the clinic, it was possible to do only two biopsies within the first 6 months. The liver biopsies were fixed in 10% formalin, processed for paraffin section. The sections were stained with hematoxylin and eosin and for reticulin fibers. Gastrocnemius muscle biopsies were performed on all 20 children initially and repeated after 6 to 8 months in eight.

Results

None of the 20 children displayed frank jaundice at any stage, although a subicteric tinge was noted in 13 cases. All the chil-

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dren, with the exception of four cases in whom the liver was not palpable initially, had soft hepatomegaly typical of protein malnutrition. This increased after 2 months and the consistency became firm. At the end of 1 year, 12 children had gross hepatomegaly and in 3 it was less, but the edges of the liver were sharp and feathery. The normal child in the series who consumed the toxic material for 1 month before she reported at the clinic still has characteristic but moderate hepatomegaly at this date (7 years later). Liver biopsy was performed after 1 month but was consistently refused later. In the child with nephrosis who consumed the toxic material for 5 days only, hepatomegaly had regressed to a just palpable liver with a sharp border after 2 years follow-up, as was also the case with another child who had consumed the toxic meal for 1 week. Three children died after 1.5 years in their respective villages after having been in hepatic coma.

After 2 years, six children, including a pair of siblings, still have enlarged firm livers (3 to 5 cm). Liver biopsies were refused by the parents. Of the remaining eight cases, there is no follow-up at this date.

Histology of the liver

The first liver biopsy was taken approximately 1 to 2 months after consumption of the toxic meal, when the majority of the children reported to the clinic. It revealed moderate to severe fatty infiltration without any changes in the lobular architecture and the presence of an inflammatory exudate extending into the hepatic parenchyma (Fig. 1). In four cases, liver biopsies showed severe fatty infiltration and an entirely undisturbed lobular pattern. In two cases, hepatic cells were severely damaged and often showed no nuclear staining with ballooning of cells. Focal necrosis and inflammatory reaction were frequently seen. Biopsy taken after 4 months

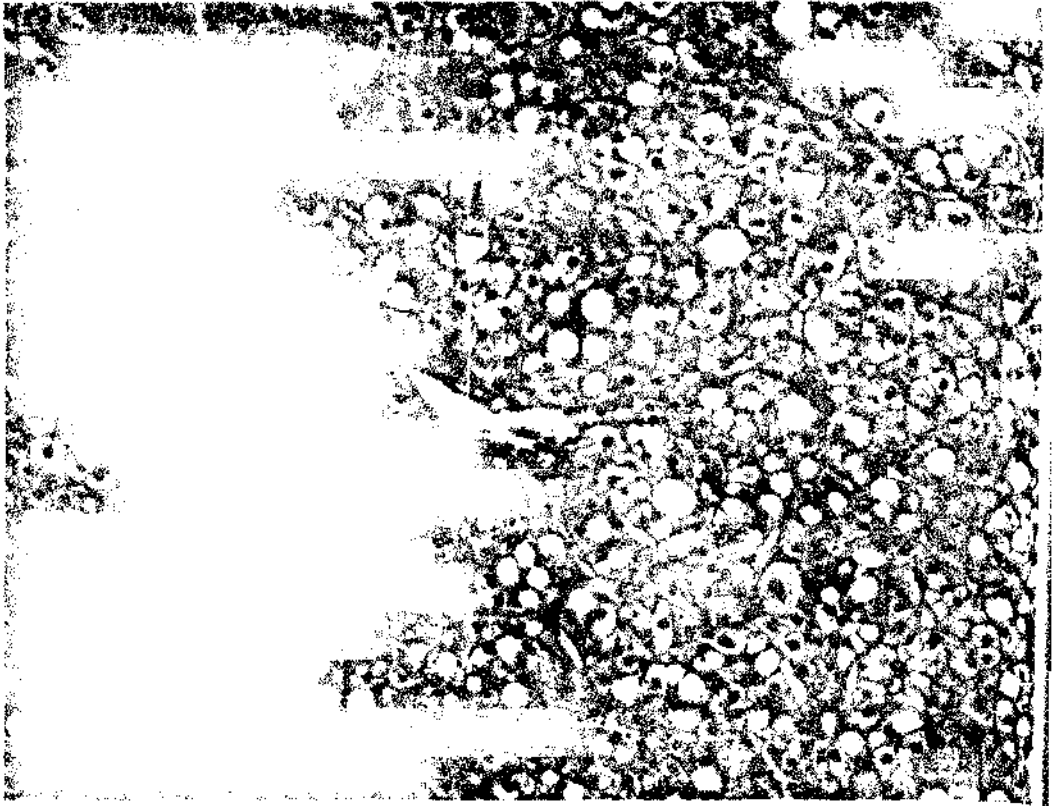


FIG. 1. Shows severe fatty infiltration of the liver. H&E \times 200.



FIG. 2. Shows strand-like inflammatory exudate associated with fibrosis of liver. H&E $\times 200$.

ned fibrous tissue appearing from the portal tract and infiltrating into surrounding parenchyma followed by severe cellular infiltration in which perlobular fibrosis was frequently seen. In two cases, septa extended into the parenchyma, but the lobule was not affected. Focal necrosis was frequently seen. The portal tract often contained inflammatory cells. Fine connective tissue membranes were seen extending throughout the parenchyma. In some areas, the connective tissue membranes were seen extending throughout the parenchyma. Also in some areas, the connective tissue membranes condensed to form septa, producing fibrosis with strand-like inflammatory exudate (Fig. 2). Biopsies taken from children between 10 months and 1 year revealed a complete abolition of the lobular architecture with dissection of septa of varying thicknesses. The typical regenerative nodules were not seen. In some cases, the hepatic cells appeared unaltered and showed no basophilia of the persisting cytoplasm. In many cases, focal coagulation occurred in the portal tract containing cells. Focal necrosis was frequently observed and bile duct proliferation

was increased. The fibrous tissue septum surrounding the lobule contained variable numbers of inflammatory cells with moderate ductular proliferation and fat-containing hepatic cells (Fig. 3). The section stained for reticulin fibers showed dissection of lobular architecture by septums of varying thicknesses (Fig. 4). Muscle biopsies done on all cases after 1 to 2 months revealed acute fragmentation with loss of striations and cellular infiltration. In the repeat biopsies done after 6 to 8 months, fragmented muscle bundles with interstitial fibrosis, fatty infiltration, and chronic inflammatory cells were seen (Fig. 5).

Discussion

The fact that kwashiorkor rarely progresses to cirrhosis is well documented (8, 9). The etiology of Indian childhood cirrhosis is obscure (4). Since 1961, 3,500 children with protein-calorie malnutrition have been treated by us on an outpatient and inpatient basis with protein-rich peanut meal. In more than 70% the follow-up has been for more

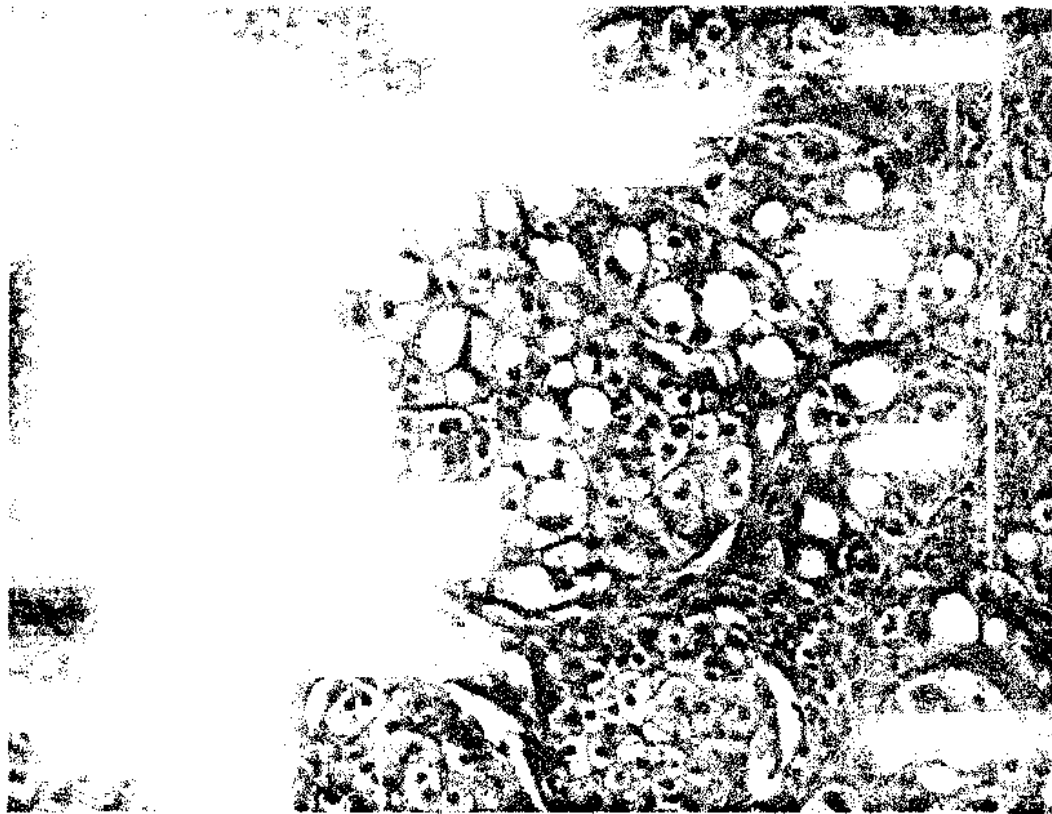


Fig. 3. Shows fibrous tissue septum surrounding the lobule. H&E. $\times 200$.

than 1 year and in several cases for more than 3 years. It was customary to perform a liver biopsy on admission, then 1 month after disappearance of edema with subsequent weight gain, and when the albumin globulin reversal came to normal. We found that fatty metamorphosis of the liver rapidly yielded to dietary protein supplements and the livers became normal within 6 weeks. In no case has cirrhosis of the liver been noted clinically or on liver biopsy material. We present this observation as a control factor in the absence of a controlled study. In contrast, the children who consumed peanut protein contaminated with aflatoxin showed a gradual transition from an increased central and periportal fat of the liver to formation of fatty cysts to fibrosis and cirrhosis over a period of 1 year. Repeated analyses of these protein-rich foods over the years reveal the aflatoxin content to be less than 30

μg per kg. The urine of cases reported here were consistently negative for aflatoxin. In our animal experiments, it was noted that aflatoxin B was excreted in the urine in small amounts within 24 hr. In these 20 children who consumed contaminated meal, the earliest signs (as is usual with Indian childhood cirrhosis also) were gastrointestinal, at which stage the toxic meal was discontinued. Urine was collected for aflatoxin content after a fortnight, when increased hepatomegaly was obvious, and the toxic meal was analyzed. We could in no way correlate the severity of liver pathology with aflatoxin found in urine of 7% of the 250 cirrhotic children studied (5). We have also found aflatoxin in the urine of 10% of normal children (5). The explanation we put forward on the basis of our experience with these kwashiorkor cases is that the exact period of toxic injury is long past by the time characteristic hepatic




FIG. 4. Shows dissection of lobular architecture by septa; reticulum stain $\times 200$.



FIG. 5. H&E section showing fibrous bands and fibrils; H&E $\times 200$.

usually occurs, and when the diagnosis of Indian childhood cirrhosis can be made. In these children, it took only 1 to 1 1/2 year for characteristic clinical and histopathological lesions to occur in the liver. The presence of aflatoxin in the urine of cirrhotic and non-cirrhotic children only confirms the on-and-on consumption of toxin containing materials (peanut oil and parboiled rice) by these children. Children with fatty liver seem to be more susceptible to toxic effects. It may be possible that both dietary and toxic components like aflatoxin are involved, as Madhavan et al. (10) have reported that protein depletion in weanling rats augmented the toxic effects of aflatoxin on the liver. The histological pictures of the livers of the cases reported here are identical to those of Indian childhood cirrhosis (4). The histopathological changes noted by us may be directly correlated with the duration of toxin ingestion. The child with nephrosis and another child who had ingested the toxic material for less than a week had mild histopathological changes that regressed considerably over a period of 1 year. The normal child and others who had consumed the contaminated meal for 1 month displayed moderately severe histological changes that were found progressive in the cases in whom serial biopsies were possible. It is of interest that characteristic skeletal muscle changes noted by us in cirrhotic children (11) and in toxin-fed rats (unpublished data) were also noted in the children who consumed the toxic meal. Some attention may be given to the possible hazards to human health from the consumption of aflatoxin-contaminated food. Some of these children are still under observation, and liver biopsies will be taken from time to time to study the progression of the lesion. These results will be published at a later date.

Summary

Childhood cirrhosis develops in varying degrees of protein deficiency and nutrition and accidentally contaminated, low fat, contaminated, and aflatoxin-contaminated, low fat, contaminated, and aflatoxin-contaminated protein from rice products ranging from 5 days to 4 weeks. The hepatic lesions showed a gradual transition from fatty liver to centrilobular and periportal cholestasis and cirrhosis which does not readily occur in treated (washed) rice. The lesions are identical to Indian childhood cirrhosis. 

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